for the Detection of Antibodies to HIV 1 and 2 in Human Serum / Plasma / Whole Blood

## INTENDED USE

The TRUSTline HIV 1/2 Ab Rapid Test is intended for use by healthcare professionals and is a rapid, qualitative, screening, lateral flow immunoassay for the simultaneous detection and differentiation of HIV-1 and HIV-2 antibodies ( $\operatorname{lgG}, \lg \mathrm{M}, \lg$ ) in human serum, plasma or whole blood. The test kit is not automated and does not require any additional instrument. Any reactive specimen with the TRUSTline HIV 1/2 Ab Rapid Test must be confirmed with alternative testing method(s) such as ELISA, Western Blot assay or PCR.

## SUMMARY AND EXPLANATION OF THE TEST

Human immunodeficiency virus type I and type II (HIV-1 and HIV-2) are enveloped, singlestranded, positive-sense RNA viruses. The causative relationship between HIV-1 and HIV-2 virus and acquired immunodeficiency syndrome (AIDS) has been established over several decades. HIV-1 has been isolated from patients with AIDS and AIDS-related complex and from healthy individuals with a high risk for developing AIDS ${ }^{1}$. HIV-2 has been isolated from West African AIDS patients and from sero-positive asymptomatic individuals ${ }^{2}$.
The two types of HIV have significant variation in sequences. HIV-1 has been divided into three groups: group M (for major) including at least ten subtypes (A through J); group O (for outlier); and group N (for non-M, non-O). Similarly, HIV-2 has been classified into at least five subtypes (A through E). Some HIV-1 variants share up to $50 \%$ homology in their envelope genes with the sequences of more common prototype strains.

Both HIV-1 and HIV-2 can elicit strong immune responses including the production of antivirus antibodies ${ }^{3}$. Presence of specific anti-HIV-1 and/or anti-HIV-2 in blood, serum or plasma indicates exposure of an individual to HIV-1 and/or HIV-2 and thus is of great value for clinical diagnosis ${ }^{4}$

The TRUSTline HIV 1/2 Ab Rapid Test was developed to detect and differentiate anti-HIV-1 and anti-HIV-2 (IgG, $\operatorname{IgM}, \operatorname{Ig} A)$ in serum, plasma or whole blood. The test can be performed by minimally trained personnel and without cumbersome laboratory equipment.

## TEST PRINCIPLE



The TRUSTline HIV 1/2 Ab Rapid Test is a lateral flow chromatographic immunoassay. The test cassette consists of: 1) a burgundy colored conjugate pad containing recombinant HIV-1 antigen conjugated with colloidal gold (HIV-1 conjugates), recombinant HIV-2 antigen conjugated with colloidal gold (HIV-2 conjugates) and a control antibody conjugated with colloidal gold, 2) a nitrocellulose membrane strip containing two test lines (1 and 2) and a control line (C). Test line 1 is pre-coated with HIV-1 antigen for the detection of antibodies to HIV-1, test line 2 is pre-coated with HIV-2 antigen for the detection of antibodies to HIV-2, and the C line is pre-coated with a control line antibody.

When an adequate volume of test specimen is dispensed into the sample well of the test cassette, the specimen migrates by capillary action across the strip. HIV-1 antibodies, if present in the specimen, migrate through the conjugate pad where they bind to the HIV-1 conjugates. The immune-complex is then captured on the membrane by the pre-coated HIV-1 antigen forming a burgundy colored line at test line 1, indicating a HIV-1 antibody positive or reactive test result. Lack of color development on test line 1 suggests an HIV-1 antibody negative or non-reactive result.
HIV-2 antibodies, if present in the specimen, migrate through the conjugate pad where they bind to the HIV-2 conjugates. The immune-complex is then captured on the membrane by the pre-coated HIV-2 antigen forming a burgundy colored line at test line 2, indicating a HIV-2 antibody positive or reactive test result. Lack of color development on test line 2 suggests a HIV-2 antibody negative or non-reactive result.

The test contains an internal control (C line), which should exhibit a burgundy colored line of the immunocomplex of the control antibodies regardless of color development on the test lines. If the C line does not develop, the test result is invalid and the specimen must be retested with another device

## REAGENTS AND MATERIALS PROVIDED

1. Individually sealed foil pouches containing
a. One cassette device
b. One desiccant

Specimen transfer device
Sample diluent ( $5 \mathrm{~mL} /$ bottle)
Package insert (instruction for use)

## MATERIALS REQUIRED BUT NOT PROVIDED

Clock or Timer
3. Alcohol swab

Lancing device for whole blood tes
4. Disposable gloves

## WARNINGS AND PRECAUTIONS

For in Vitro Diagnostic Use

1. This package insert must be read completely before performing the test. Failure to follow the insert may lead to inaccurate test results.
Do not open the sealed pouch unless ready to conduct the assay
Do not use the test device if pouch is not intact.
Do not use expired devices or components.
Bring all reagents to room temperature $\left(15-30^{\circ} \mathrm{C}\right)$ before use
2. Do not use the components of different lots and of any other type of test kit as a substitute for the components in this kit.
3. Do not use hemolyzed blood for testing
4. Wear protective clothing and disposable gloves while handling the kit reagents and clinical specimens. Wash hands thoroughly after performing the test.
Users of this test should follow the US CDC Universal Precautions for prevention of transmission of HIV, HBV and other blood-borne pathogens.
Do not smoke, drink or eat in areas where specimens or kit reagents are being handled.
5. Dispose of all specimens and materials used to perform the test as bio-hazardous waste
6. Handle the negative and positive controls in the same manner as the patient specimens.
7. The test result should be read 15 minutes after a specimen is applied to the sample well of the device. Reading the test result after 20 minutes may give erroneous results.
8. Do not perform the test in a room with strong air flow, i.e. an electric fan or strong air conditioning.
9. Clean up spills thoroughly using appropriate disinfectant.

## REAGENT PREPARATION AND STORAGE INSTRUCTIONS

All reagents are ready to use as supplied. Store test kit at $1-30^{\circ} \mathrm{C}$. If stored at $2-8^{\circ} \mathrm{C}$, ensure that all reagents are brought to room temperature before opening. The sample diluen (opened and unopened) and unopened test device is stable through the expiration date printed on the label, when stored at recommended temperature. Do not freeze the kit or expose the kit to temperatures above $30^{\circ} \mathrm{C}$. The test device is sensitive to humidity and heat. Perform the test immediately after removing the test device from the foil pouch

## SPECIMEN COLLECTION AND HANDLING

Consider any materials of human origin as infectious and handle them using standard biosafety procedures

Plasma
Step 1: Collect blood specimen into a lavender, blue or green top collection tube (containing EDTA, citrate or heparin, respectively, in Vacutainer®) by venipuncture.
Step 2: Separate the plasma by centrifugation
Step 3: Carefully withdraw the plasma into a new pre-labeled tube.
Serum
Step 1: Collect blood specimen into a red top collection tube (containing no anticoagulants in Vacutainer ${ }^{\circledR}$ ) by venipuncture.
Step 2: Allow the blood to clot.
Step 3: Separate the serum by centrifugation.
Step 4: Carefully withdraw the serum into a new pre-labeled tube.
Test specimens as soon as possible after collecting. Store specimens at $2-8^{\circ} \mathrm{C}$, if not tested immediately. The specimens can be stored at $2-8^{\circ} \mathrm{C}$ for up to 5 days. The specimens should be frozen at $-20^{\circ} \mathrm{C}$ for longer storage.

Avoid multiple freeze-thaw cycles. Prior to testing, bring frozen specimens to room temperature slowly and mix gently. Specimens containing visible particulate matter should be clarified by centrifugation before testing.

Do not use samples demonstrating gross lipemia, gross hemolysis or turbidity in order to avoid interference with result interpretation.

Blood
Drops of whole blood can be obtained by either finger tip puncture or venipuncture. Collect blood specimen into a lavender, blue or green top collection tube (containing EDTA, citrate or heparin, respectively, in Vacutainer®). Do not use hemolyzed blood for testing. Capillary blood (finger tip puncture) can be used directly without anticoagulant. Collect blood with Specimen transfer device and transfer it to sample well of device

Whole blood specimens should be stored in refrigeration $\left(2-8^{\circ} \mathrm{C}\right)$, if not tested immediately The specimens must be tested within 24 hours of collection

## ASSAY PROCEDURE

Step 1: Bring the specimen and test components to room temperature if refrigerated or frozen. Once thawed, mix the specimen well prior to performing the assay.

Step 2: When ready to test, open the pouch at the notch and remove device. Place the test device on a clean, flat surface.

Step 3: Be sure to label the device with specimen's ID number.
Step 4: Fill the specimen transfer device with specimen (about $20 \mu \mathrm{~L}$ ) not to exceed the specimen line as shown in the images below. For better precision, transfer specimen using a pipette capable of delivering a $20 \mu \mathrm{~L}$ volume.

Holding the specimen transfer device vertically, dispense the entire specimen into the center of the sample well making sure that there are no air bubbles.

Immediately add 2 drops $(60-80 \mu \mathrm{~L})$ of sample diluent to the sample well with the bottle positioned vertically

$20 \mu \mathrm{~L}$ specimen

$20 \mu \mathrm{~L}$ specimen


Result at 15 minutes

2 drops of sample diluent $\rangle$


2 drops of sample diluent

Step 5: Set up timer
Step 6: Result should be read at 15 minutes.
Do not read result after 20 minutes. To avoid confusion, discard the test device after interpreting the result.

## QUALITY CONTROL

1. Internal Control: This test contains a built-in control feature, the $C$ line. The $C$ line develops after adding specimen and sample diluent. If the $C$ line does not develop, review the entire procedure and repeat the test with a new device.
2. External Control: Good Laboratory Practice recommends using external controls, positive and negative, to assure the proper performance of the assay, particularly under the following circumstances:
a. A new operator uses the kit, prior to performing testing of specimens
b. A new lot of test kits is used.
c. A new shipment of test kits is used.
d. The temperature used during storage of the kit falls outside of $1-30^{\circ} \mathrm{C}$
e. The temperature of the test area falls outside of $15-30^{\circ} \mathrm{C}$.
f. To verify a higher than expected frequency of positive or negative results.
g. To investigate the cause of repeated invalid results.

## INTERPRETATION OF ASSAY RESULT

1. NEGATIVE RESULT

If only the C line is developed, the test indicates that the level of anti-HIV-1 and anti-HIV-2 in the specimen is undetectable. The result is negative or non-reactive for HIV1 and negative or non-reactive for HIV-2.
2. POSITIVE RESULT:
2.1 If both the $C$ line and test line 1 are developed, the test indicates that the specimen contains anti-HIV-1. The result is HIV-1 positive or reactive.

2.3 If the C line and both test lines ( 1 and 2 ) are developed, the test indicates that the specimen contains anti-HIV-1 and anti-HIV-2. The result is HIV-1 positive or reactive and HIV-2 positive or reactive. For differentiation of the type of HIV virus infection, see Limitations of Test section, Number 5.


Samples with reactive results should be confirmed with alternative testing method(s) such as Western Blot assay, PCR or ELISA and clinical findings before a final diagnostic decision is made.

## 3. INVALID:

If no C line is developed, the assay is invalid regardless of color development in the test lines as indicated below. Repeat the assay with a new device.


PERFORMANCE CHARACTERISTICS

1. Clinical Performance for HIV-1 Ab Test

A total of 1308 samples from susceptible subjects were tested with the TRUSTline HIV 1/2 Ab Rapid Test and with a commercial HIV Ab ELISA Kit. Comparison for all subjects is shown in the following table.

|  | TRUSTline HIV 1/2 Ab Rapid Test |  |  |
| :--- | :---: | :---: | :---: |
| ELISA | Positive | Negative | Total |
| Positive | 326 | 0 | 326 |
| Negative | 2 | 980 | 982 |
| Total | 328 | 980 | 1308 |

Relative Sensitivity: 100\%, Relative Specificity: 99.79\%, Overall Agreement: 99.84\%
2. Clinical Performance for HIV-2 Ab Test

A total of 195 samples from susceptible subjects were tested with the TRUSTline HIV 1/2 Ab Rapid Test and with a commercial HIV Ab ELISA Kit. Comparison for all subjects is shown in the following table.

|  | TRUSTline HIV 1/2 Ab Rapid Test |  |  |
| :--- | :---: | :---: | :---: |
| ELISA | Positive | Negative | Total |
| Positive | 49 | 0 | 49 |
| Negative | 1 | 174 | 175 |
| Total | 50 | 174 | 224 |

Relative Sensitivity: 100\%, Relative Specificity: 99.43\%, Overall Agreement: 99.55\%
3. Cross-Reactivity

The negative sample was spiked with serum specimens of infectious diseases and then tested according to the standard procedure. The results showed that the TRUSTline HIV $1 / 2$ Ab Rapid Test had no cross-reaction with the following tested serum specimens of infectious disease.

| Specimen | Sample <br> Size | HIV-1 Ab <br> Reactivity | HIV-2 Ab <br> Reactivity |
| :--- | :---: | :---: | :---: |
| HBsAg Positive Serum | 10 | Negative | Negative |
| HAV Positive Serum | 10 | Negative | Negative |
| HCV Positive Serum | 10 | Negative | Negative |
| Dengue Positive Serum | 10 | Negative | Negative |
| Syphilis Positive Serum | 10 | Negative | Negative |
| ANA Positive Serum | 5 | Negative | Negative |
| RF Positive Serum <br> $(\leq 2,500$ IU/ml) | 5 | Negative | Negative |

3. Precision

Within run and between run precisions have been determined by testing 20 replicates with four categories of the specimens: negative, weak, medium and strong positive specimens. The negative, weak, medium and strong positive specimens were correctly identified in all of the tests performed in each run.
4. Interference

Common substances (such as pain and fever medication, blood components) may affect the performance of the TRUSTline HIV 1/2 Ab Rapid Test. This was studied by spiking of these substances into the three levels of the HIV-1 Ab and HIV-2 Ab standard control. The results are presented in the following table and demonstrate that the substances studied did not affect the performance of the TRUSTline HIV 1/2 Ab Rapid Test.
Note: -: Negative; +: Weak Positive; +++: Strong Positive

| Potential Interfering Substances Spiked | HIV-1 Ab Reactivity |  |  | HIV-2 Ab Reactivity |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: |
|  | Negative | Weak Positive | Strong Positive | Negative | Weak Positive | Strong Positive |
| Control | - | + | +++ | - | + | +++ |
| Bilirubin $20 \mathrm{mg} / \mathrm{dL}$ | - | + | +++ | - | + | +++ |
| Creatinine $442 \mu \mathrm{~mol} / \mathrm{L}$ | - | + | +++ | - | + | +++ |
| Glucose $55 \mathrm{mmol} / \mathrm{L}$ | - | + | +++ | - | + | +++ |
| Albumin $60 \mathrm{~g} / \mathrm{L}$ | - | + | +++ | - | + | +++ |
| Salicylic Acid 4.34 $\mathrm{mmol} / \mathrm{L}$ | - | + | +++ | - | + | +++ |
| Heparin 3,000 U/L | - | + | +++ | - | + | +++ |
| EDTA $3.4 \mu \mathrm{~mol} / \mathrm{L}$ | - | + | +++ | - | + | +++ |

1. The Assay Procedure and Interpretation of Assay Result sections must be followed closely when testing for the presence of antibodies to HIV in serum, plasma or whole blood from individual subjects. Failure to follow the procedure may lead to inaccurate test results.
2. The TRUSTline HIV $1 / 2 \mathrm{Ab}$ Rapid Test is limited to the qualitative detection of HIV-1 or HIV-2 antibodies in human serum, plasma or whole blood. The intensity of the test line does not correlate with the antibody titer of the specimen.
3. A non-reactive result for an individual subject indicates absence of detectable HIV-1 or HIV-2 antibodies. However, a non-reactive test result does not preclude the possibility of exposure to or infection with HIV-1 or HIV-2.
4. A non-reactive result can occur if the quantity of the HIV-1 or HIV-2 antibodies present in the specimen is below the detection limits of the assay or the antibodies that are detected are not present during the stage of disease in which a sample is collected.
5. As illustrated in the Interpretation of Assay Result, Section 2.3, all three test lines (1, 2 and C) may develop when tested with samples containing high titers of HIV-1 antibodies. Hence reactive test bands for both HIV-1 and HIV-2 may not indicate mixed infection but might from the cross-reactivity of HIV-1 and HIV-2 because of the similarity of their genomic structure. To differentiate and to resolve antibody crossreactivity, dilute the test specimen with sample diluent 1:50 or 1:100, then re-test the diluted specimen with a new test device. Only test line 1 and the $C$ line will appear if the specimen contains antibodies to HIV-1. If test line 1, test line 2 and the C line all appear, the test indicates presence of antibodies to both HIV-1 and HIV-2.
6. False negative results may arise because of hook effect due to very high titer of antibody in sample. Repeat the test by using different dilution of same sample.
7. Hemolytic samples may give reddish background even after end of the test time.
8. Infection may progress rapidly. If symptoms persist while the result from the TRUSTline HIV 1/2 Ab Rapid Test is non-reactive, it is recommended to test with an alternative test method or to re-sample the patient a few days later.
9. Unusually high titers of heterophile antibodies or rheumatoid factor in specimens may affect expected results.
10. The results obtained with this test should only be interpreted in conjunction with other diagnostic procedures and clinical findings

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## Index of Symbols



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